

Altered composition of the Indonesian gut microbiome and heavy metal resistance genes abundance in response to heavy metal exposure

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Abstract: Exposure to heavy metals poses significant health risks through inhalation, ingestion, and contact with contaminants, potentially causing gastrointestinal disorders and heightened infection vulnerability by altering the gut microbiome. In Indonesia, heavy metal contamination and its impact on the gut microbiome remain underexplored. This study assessed heavy metal prevalence in stool samples from coastal and highland populations and its potential microbiome effects. Copper (Cu), barium (Ba), manganese (Mn), and zinc (Zn) were detected, with barium contamination particularly concerning (13 of 20 participants). Genes for heavy metal resistance, such as znuC, nikE, modC, mntH, and arsB, were common in both populations. Copper levels negatively correlated with Prevotella abundance, while barium levels positively correlated with Prevotella, Faecalibacterium, and Ruminococcus abundance, indicating an antagonistic Ba-Cu relationship in shaping the microbiome. This study highlights the health risks of heavy metal exposure in Indonesia and its potential impact on gut microbiome composition.

Keywords: heavy metals exposure; copper; barium; health risks; gut microbiome.

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1 Introduction

Heavy metals are elements with high atomic weights that can be toxic to humans, even in small quantities (Hordyjewska et al., 2014). They can accumulate in the human body typically through inhalation, ingestion, or skin contact with contaminated substances or environments (Wu et al., 2019). Certain foods, especially those grown in contaminated soil or exposed to contaminated air or water sources, may contain higher levels of heavy metals (Sandeep et al., 2019; Turkyilmaz et al., 2020). Seafood can also be a source of heavy metal exposure (Ziyuan et al., 2022). When heavy metals are ingested, they can be absorbed into the bloodstream through the digestive tract, leading to potential health risks (Sarpong-Kumankomah et al., 2018).

Some heavy metals can cause direct damage to the gastrointestinal tract, leading to symptoms such as abdominal pain and diarrhoea (Akpor, 2014). Heavy metals absorbed from the digestive system can also have systemic effects on the body beyond the gastrointestinal tract. They can accumulate in the liver and kidneys, which can impact their function over time (Sumino et al., 1975). They can affect the nervous system, causing cognitive impairments and behavioural changes (Sankhla et al., 2017; Sumino et al., 1975). Heavy metals exposure also increases the risk of various cancers (Lim et al., 2019; Sá et al., 2016), affects the cardiovascular system (Sevim et al., 2020), and increases the risk of diabetes (Javaid et al., 2021).

Recent work shows that heavy metals can have significant effects on the human gut microbiome. Heavy metal exposure can lead to alterations in the composition and diversity of the gut microbiota, causing shifts in the abundance and diversity of certain microbial species in the gut (Dong et al., 2017; Zhang et al., 2021, 2023). They also can also alter the expression of genes involved in various metabolic pathways, including those related to nutrient metabolism, energy production, and detoxification (Fatema et al., 2021; Gao et al., 2017). Further altered microbial composition and function due to heavy metal exposure may contribute to gastrointestinal disorders and increased susceptibility to infections (Brabec et al., 2020; Zhang et al., 2023).

Indonesia, the fourth-most populous country, also faces challenges related to heavy metal exposure due to various factors, including industrial activities and environmental pollution (Ahmad et al., 2022; Yap and Al-Mutairi, 2021). The use of certain fertilisers and pesticides containing heavy metals resulted in the accumulation of these metals in

agricultural soils (Budianta et al., 2023). Additionally, the application of sewage sludge or improperly treated wastewater for irrigation purposes introduced heavy metals into the food chain (Sudarningsih et al., 2017). Heavy metals contamination in water sources due to mining activities, industrial discharges, and improper waste disposal practices also reported (Gafur et al., 2018; Juliani, 2021; Palapa and Maramis, 2015). There are considerable reports regarding heavy metals contamination in drinking water (Astuti et al., 2021), agricultural products (Wahyuningsih et al., 2023), fresh water fish (Nasir et al., 2021) and seafood (Puspitasari et al., 2023) in Indonesia.

Despite the high risk of heavy metal exposure through water and food among Indonesians, there is limited data on the contamination rates of heavy metals in the human body within the country. While contamination of heavy metals in urine and blood samples has been reported among high-risk populations such as mining (Ekawanti and Krisnayanti, 2015) and industrial workers (Damastuti et al., 2017), there is a notable absence of data regarding heavy metal contamination in the digestive tract and in the general population. Further, the effect of heavy metal exposure to Indonesian gut microbiome is never evaluated before. This study focuses on reporting heavy metal contamination in faecal samples collected from both the coastal and highland populations of Indonesia, as well as investigating their potential effects on gut microbial composition.

2 Material and method

2.1 Study participants, stool sample collection and DNA extraction

For this study, we utilised stool samples that had been collected in previous studies (Luqman et al., 2021; Wibowo et al., 2021). These samples originated from two distinct study populations: the coastal population of Kenjeran, Surabaya, Indonesia (consisting of 7 males and 2 females), and the highland population of Pacet, Mojokerto, Indonesia (comprising 5 males and 6 females). The selection of participants was based on specific criteria, including being in a healthy condition, aged between 20 and 50 years old. The DNA extraction from the collected stool samples was conducted using the Zymbiomic DNA Miniprep Kit (Zymo Research), following the provided protocol from the company.

2.2 Normalised read counts

The analysis of gene read counts was performed using previously published methods (Luqman et al., 2021; Nugrahapraja et al., 2022). The metagenomic data used in this study was sourced from the DNA Data Bank of Japan (DDJP) with the submission number SSUB023028 (Nugrahapraja et al., 2022). The sequences corresponding to the target or reference genes (provided in Table S1) were downloaded from the NCBI database. To compare the data between samples and genes, all reads from the samples were aligned against these sequences using BWA (Li et al., 2019), and the resulting samfiles were processed with SAM tools to extract the number of mapped reads (Li et al., 2009). The read counts were subsequently normalised by taking into account the number

of sequenced reads and the length of the respective genes, ensuring comparability across samples and genes.

2.3 Detection and measurement of heavy metals from faecal samples

As much as 10 g of faecal samples was analysed using X-Ray Fluorescence NitonTM XL2 GOLDD (Thermo Scientific, Carlsbad, CA, USA) to calculate heavy metals content in the samples. The heavy metals measurements were presented as percentages of the weight of the faeces.

2.4 Data analysis

Comparation of heavy metals concentration in stool samples collected from coastal and highland populations of Indonesia were analysed using t-test. In addition, to determine possible connection between heavy metal concentration and heavy metal related-genes abundance, and also connection between heavy metal concentration and bacterial genus abundance in both population (highland population and coastal population) were analysed using Spearman's correlation.

3 Results

3.1 Detection of heavy metals in coastal and highland faecal samples

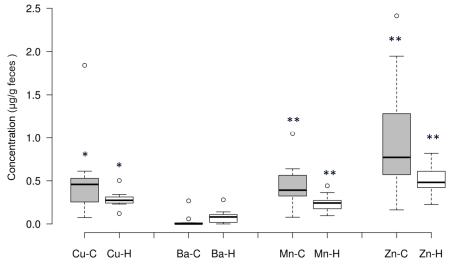
Stool samples were collected from a total of 20 participants, with 9 individuals (7 males and 2 females) representing a fisherman community residing in the coastal region of Surabaya, Indonesia, and the remaining 11 participants (5 males and 6 females) belonging to a farmer community residing in the highland region of Pacet, Indonesia. Heavy metal analysis revealed the presence of four types of heavy metals, namely copper (Cu), barium (Ba), manganese (Mn), and zinc (Zn) in the study population. Cu, Mn, and Zn were detected in all samples, while Ba was found in 13 participants (Table S2). The average concentrations of Cu, Ba, Mn, and Zn in the coastal samples were 0.534 μ g/g, 0.037 μ g/g, 0.454 μ g/g, and 1.01 μ g/g respectively. In the highland samples, the average concentrations of Cu, Ba, Mn, and Zn were 0.282 μ g/g, 0.082 μ g/g, 0.240 μ g/g, and 0.509 μ g/g respectively (Figure 1). These findings indicate that heavy metal was observed in the digestive tracts of all study participants, with higher levels of Cu, Mn, and Zn detected in the coastal population, while Ba contamination was more pronounced in the highland population.

3.2 Heavy metals related-genes are widely spread in Indonesians gut microbiome

We conducted an analysis of the relative gene abundance using previously collected stool samples from coastal and highland populations in Indonesia. The samples were mapped against a reference set of heavy metal-related genes (Table S1) known to be involved in the response and metabolism of heavy metals in living organisms. These genes play a crucial role in the uptake, transport, detoxification, and excretion of heavy metals from cells and tissues (Pal et al., 2014). They are responsible for regulating various cellular

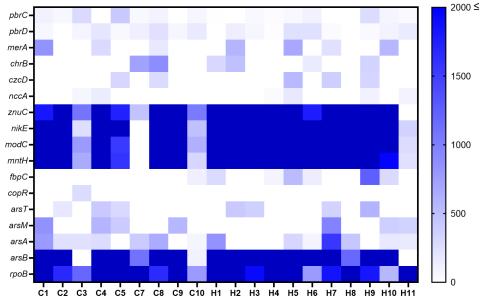
processes and pathways that enable organisms to cope with heavy metal exposure (Xavier et al., 2019).

Figure 1 Concentration of heavy metals in stool samples collected from coastal and highland populations of Indonesia



Abbreviations of samples: C – Coastal, H – Highland, $*P \le 0.1$, $**P \le 0.05$ on t-test.

Figure 2 The prevalence and abundance of heavy metal related genes in Indonesian gut microbiota (A) the heatmap of the relative reads of the heavy metal related genes with RpoB (housekeeping gene) as a comparison (see online version for colours)



Abbreviations of samples: C - Coastal, H - Highland.

Our analysis revealed that the heavy metal-related genes were widespread in the stool samples from both populations (Figure 2). Certain genes, such as znuC, nikE, modC, and mntH that regulate zinc, nickel, molybdenum, and manganese transport, respectively, were prevalent and detected in 19 out of 20 stool samples, while arsB that regulate arsenic resistance was detected in 18 out of 20 samples (Table S3). These genes also exhibited high relative abundance in the stool samples, indicating their strong and widespread distribution in the human gut. On the other hand, the gene copR, which is involved in the regulation of copper resistance mechanisms, was only detected in one sample and showed relatively low abundance (Figure 2 and Table S3).

3.3 The effect of heavy metal contamination on heavy metal related-genes abundance

The expression and activity of heavy metals related-genes can be influenced by exposure to heavy metals (Das et al., 2016). Understanding their functions and regulation is important for studying the mechanisms of heavy metal toxicity, as well as for developing strategies to mitigate the adverse effects of heavy metal exposure in biological systems. Therefore, we performed spearman correlation analysis to determine correlations between heavy metal contamination rate with heavy metal related-genes abundance.

Table 1	Correlation between heavy metal concentration and heavy metal related-genes
	abundance

Heavy metal related genes	Significantly correlated metals	Correlation coefficient	P-value
modC	Ba	0.562	0.009913
znuC	Ba	0.552	0.01161
mntH	Cu	-0.4719	0.03926
	Ba	0.5527	0.0115
nikE	Cu	-0.4647	0.04231
	Ba	0.4822	0.03132
merA	Cu	-0.4552	0.04374
	Ba	0.4737	0.03487

Our findings revealed a notable positive correlation between barium (Ba) levels and the abundance of specific heavy metal-related genes (Table 1). Notably, genes such as modC, znuC, mntH, nikE, and merA displayed a significant positive correlation with barium levels observed in the stool samples. These results suggest that elevated concentrations of Ba may impact the abundance of these genes, which are involved in the response and metabolism of heavy metals. Conversely, copper (Cu) levels appeared to have a negative influence on the abundance of mntH, nikE, and merA genes (Table 1). This indicates that higher concentrations of Cu may contribute to reduced abundance of these particular heavy metal-related genes. Overall, our study demonstrates that heavy metal exposure, particularly in relation to Ba and Cu can significantly affect the abundance of heavy metal-related genes within the human gut. These findings shed light on the

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potential impact of heavy metal exposure on gene regulation and highlight the complex relationship between heavy metals and gene abundance in the context of gut health.

3.4 The effect of heavy metal contamination on gut microbial composition

Previous research has reported substantial impact of heavy metal contamination on the composition of the human gut microbiome. Heavy metal exposure can lead to a decrease in microbial diversity, causing shifts in the relative abundance of different bacterial taxa. Some studies have shown that heavy metals can selectively promote the growth of certain metal-resistant bacteria while suppressing the growth of other beneficial microbial species (Brabec et al., 2020; Zhang et al., 2023). To investigate the influence of heavy metal exposure on the gut microbiome of the Indonesian population, we conducted Spearman correlation analysis to assess the relationship between heavy metal levels and the relative abundance of microbial species in two distinct study populations.

In the highland population, we observed a robust negative correlation between Cuprum (Cu) levels and the abundance of Prevotella (Table 2), indicating that Cu has a detrimental effect on the growth and propagation of Prevotella. Furthermore, we found a strong negative correlation between Barium (Ba) and Escherichia, suggesting that Ba may limit the growth of Escherichia in the gut. Interestingly, we observed a positive correlation between Ba levels and the abundance of Faecalibacterium and Ruminococcus, implying that these bacterial genera can thrive in an environment with elevated Ba levels. Similarly, we noted a positive correlation between manganese (Mn) levels and the abundance of Ruminococcus and Clostridium. This suggests that these two bacterial genera are susceptible to higher Mn levels and may exhibit increased abundance in the presence of Mn.

Table 2 Correlation between heavy metal concentration and bacterial genus abundance in highland population

	Significantly correlated		
Bacterial genus	metals	Correlation coefficient	P-value
Prevotella	Ba	0.722	0.02806

In the coastal population, we observed a significant positive correlation between Barium (Ba) levels and the abundance of Prevotella (Table 3), suggesting an antagonistic relationship between Ba and Cu in influencing Prevotella abundance. Specifically, Ba appeared to promote the growth of Prevotella, while Cu seemed to have a limiting effect on its abundance. Overall, these findings shed light on the complex interactions between heavy metals and specific bacterial genera in the gut microbiome. The observed correlations provide insights into how heavy metal exposure can influence the growth and distribution of particular microbial species, thereby impacting the overall composition and functioning of the human gut microbiome.

Table 3 Correlation between heavy metal concentration and bacterial genus abundance in coastal population

Bacterial genus	Significantly correlated metals	Correlation coefficient	P value
Prevotella	Cu	-0.7973	0.003292
Faecalibacterium	Mn	0.6821	0.02077
	Ba	0.6929	0.01809
Ruminococcus	Ba	0.8904	0.0002383
	Mn	0.6228	0.04068
Escherichia	Ba	-0.7715	0.005417
Clostridium	Mn	0.6844	0.02018

4 Discussion

Indonesia, like many other countries, faces health problems related to heavy metal pollution (Ahmad et al., 2022; Yap and Al-Mutairi, 2021), which arises from various sources such as industrial activities (Palapa et al., 2015; Juliani et al., 2021), improper waste disposal (Sudarningsih et al., 2017), and agricultural practices (Budianta et al., 2023). These contaminants can enter the environment, contaminate water sources (Astuti et al., 2021), soil (Budianta et al., 2023), and food (Puspitasari et al., 2023; Wahyuningsih et al., 2023), and eventually affect human health. Despite the evident hazards, there is currently a lack of data regarding heavy metal contamination in faecal samples from the general population in Indonesia and. To address this knowledge gap, our study focuses on examining the prevalence of heavy metal exposure among Indonesia's coastal and highland populations, while also evaluating their impact on the composition of the gut microbiome.

In our analysis, we did not detect the presence of highly toxic heavy metals such as lead, mercury, cadmium, arsenic, and chromium in the samples, indicating that contamination with these highly toxic metals is rare among the general population. However, we did identify the presence of other heavy metals including copper (Cu), barium (Ba), manganese (Mn), and zinc (Zn) in the samples. Copper, manganese, and zinc is an essential nutrient for living organisms, playing a vital role in enzyme function and metabolism of proteins and carbohydrates (Pajarillo et al., 2021). While copper, manganese, and zinc are necessary for human health in small amounts, excessive exposure to these metals can lead to health issues (Chasapis et al., 2020; Taylor et al., 2020). The average concentrations of copper (Cu), manganese (Mn), and zinc (Zn) in the coastal samples were measured at 0.534 µg/g, 0.454 µg/g, and 1.01 µg/g, respectively. In the highland samples, the average concentrations of Cu, Mn, and Zn were found to be $0.282 \mu g/g$, $0.240 \mu g/g$, and $0.509 \mu g/g$, respectively. These concentrations fall within the normal range typically observed in healthy adult humans (Bost et al., 2016; Hordyjewska et al., 2014). Therefore, it can be inferred that the study population does not exhibit any significant metal intoxication issues related to these three metals.

Barium (Ba) exposure in humans has garnered attention due to its increasing industrial and medicinal utilisation over the past four decades. Although Ba has always been present as a trace element in food and drinking water, its prevalence has significantly risen (Kravchenko et al., 2014; Peana et al., 2021). Our study revealed a high prevalence of barium contamination among the participants, with 13 out of 20 individuals affected. This finding is worrisome as barium exposure has been associated with various health concerns, including elevated blood pressure (Zeng et al., 2022), gastrointestinal disturbances and muscle weakness (Bhoelan et al., 2014), kidneys and cardiovascular malfunction (Dallas and Williams, 2001), neurological impairments (Wang et al., 2021), and mental disorders. Further investigations are imperative to fully understand the extent of barium exposure in the general Indonesian population and identify potential sources of contamination, particularly amongst vulnerable groups such as the elderly, pregnant women, and children.

To evaluate the impact of heavy metal on the human gut microbiome, we first investigate their effect on the abundance of heavy metal related-genes. We found that some genes such as znuC, nikE, modC, mntH, and ArsB are highly prevalent and abundant in the human gut. These genes regulate the uptake and efflux of zinc (Kandari et al., 2021), nickel (Higgins, 2019), molybdenum (Huang et al., 2022), manganes (Kajfasz et al., 2020), and arsenic (Shen et al., 2013), respectively, in various bacterial species. The abundance of these genes in the human gut suggest the selective pressure of metals concentration in the human gut that favour the growth of metal resistance bacteria. Consistent with these findings, previous studies have reported higher abundance and expression of ArsB in children with high arsenic exposure compared to matched controls (Dong et al., 2017). Further, we found that the abundance of metal resistant genes is significantly correlated with Ba dan Cu concentration in faecal samples, where Ba has a strong positive effect on gene abundance while Cu shows the opposite effect. This result indicates that high Ba contamination in the gut could lead to the selection of bacteria that carry metal resistance genes, thus affecting the composition of gut microbiome. In accordance, several works have reported that various heavy metal pollution could induce the emergence and enrichment of metal resistant bacteria (Mustapha and Halimoon, 2015; Zampieri et al., 2016).

Consistent with these findings, our study revealed a positive correlation between barium (Ba) concentration in faecal samples and the abundance of several bacterial genera, including Faecalibacterium, Ruminococcus, and Prevotella. Conversely, we observed a strong negative correlation between Ba concentration and the abundance of Escherichia. These results align with previous research indicating that exposure to heavy metals such as arsenic and cadmium can have a positive effect on Prevotella (Li and Durbin, 2009) and Faecalibacterium abundance but a negative effect on Escherichia. reported an association between arsenic exposure and the depletion of specific gut commensal bacteria Ruminococcus (Brabec et al., 2020). In contrast, our findings revealed a positive correlation between barium exposure and the abundance of Ruminococcus. These results indicate that different metals can have distinct effects on specific bacteria, which may also be influenced by the physiological state of the host. Taken together, our findings support the notion that heavy metal exposure can selectively promote the growth of certain bacteria, potentially those with metal-resistant properties, while suppressing the growth of other microbial species. This suggests that heavy metals

may exert a significant influence on the composition and dynamics of the gut microbiome, favouring the proliferation of metal-tolerant bacteria. Further research is warranted to elucidate the underlying mechanisms and explore the broader implications of these findings on human health and microbial ecology.

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Author contributions

Conceptualization, A.L. and A.T.W.; methodology, A.L., A.T.W, F.N.H; validation, A.L., A.T.W., and R.A.W.; formal analysis, A.L., F.N.H., and A.T.W; investigation, F.N.H and M.A.K..; resources, A.L., A.T.W., and R.A.W.; data curation, A.L., F.N.H., M.A.K., and A.T.W; writing—original draft preparation, A.L. and A.T.W.; writing—review and editing, A.L and A.T.W.; visualization, A.L. and A.T.W.; supervision, A.L.; project administration, A.L., AT.W., and R.A.W..; funding acquisition, A.L. and A.T.W. All authors have read and agreed to the published version of the manuscript.

Ethical approval

The collection of human stool samples was approved by the Health Research Ethic Committee of Universitas Surabaya (No. 005-OL/KE/III/2021). Stool samples were obtained from 11 adult participants from the highland population of Pacet, Mojokerto, Indonesia and 11 adult participants from the coastal population of Kenjeran, Surabaya, Indonesia. All samples were anonymized and obtained with written consent from the participants.

Declaration of competing interest

The authors declare no conflict of interest.

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Supplementary material

 Table S1
 Reference genes

No.	Gene	Reference species	Function	Accession number	Locus tag
1	гроВ	Bacteroides intestinalis	DNA-directed RNA polymerase subunit beta	NZ_QRKQ01000009.1	DW169_RS10850
2	merA	S. aureus	Mercury reductase (II)	NC_013352.1	HHF46_RS00110
3	chrB	Ralstonia solanacearum	Chromate resistance protein	NC_021745.1	F504_3817
4	czcD	Caulobacter vibrioides	cobalt-zinc-cadmium resistance protein	NC_011916.1	CCNA_00305
5	nccA	Parachlamydia acanthamoebae	nickel-cobalt-cadmium resistance protein	NC_015702.1	PUV_25170
6	znuC	E. coli	Zn(2(+)) ABC transporter ATP binding subunit	NC_000913.3	ECK1859
7	nikE	E. coli	Ni(2(+)) ABC transporter ATP binding subunit	NC_000913.3	ECK3464
8	modC	E. coli	molybdate ABC transporter ATP binding subunit	NC_000913.3	ECK0754
9	mntH	E. coli	Mn(2(+))/Fe(2(+)): H(+) symporter	NC_000913.3	ECK2387
10	fbpC	Neisseria meningitidis	iron(III)-transport system ATP-binding	NC_017512.1	NMAA_0482
11	copR	Salmonella enterica	Copper resistance transcriptional regulatory protein	NC_003197.2	STM1096
12	arsT	Corynebacterium urealyticum	putative arsenic-transport protein	NC_020230.1	CU7111_0032
13	arsM	Thermacetogenium phaeum	S-adenosyl-L- methionine:arsenic(III) methyltransferase	NC_018870.1	Tph_c04450
14	arsA	Enterobacter cloacae	arsenite efflux transporter ATPase subunit		EC036_RS04535
15	arsB	E. coli	arsenite/antimonite:H(+) antiporter		b3502, ECK3487, arsF
16	pbrD	Cupriavidus metallidurans CH34	Pb(II) binding protein	NC_007971.2	Rmet_5949
17	pbrC	Cupriavidus metallidurans CH34	lead resistance fusion protein	NC_006466.1	pMOL30_090

 Table S2
 Concentration of heavy metals in stool samples collected from coastal and highland populations of Indonesia

Heavy metals concentration											
(mg/g)					Coasta	Coastal population samples	samples				
	CI	C2	C3	C4	C5	C7	C8	6.3	CI0		
Cu	0.242556	0.499106	0.499106 0.611812		0.458076	0.298659 0.458076 0.528025 1.840979	1.840979	0.074127	0.25441		
Mn	0.225828	0.472962	0.472962 0.638413	0.323547	0.391447	0.391447 0.563227 1.047454	1.047454	0.077834	0.352261		
Zn	0.602208	1.283414	1.283414 2.412668		0.978617	0.771536 0.978617 1.946653 0.571338	0.571338	0.16308	0.410971		
Ba	0.267648	0	0	0.012444	0	0	0	0	0.05871		
					Highlar	Highland population samples	1 samples				
	IHI	H2	H3	H4	H5	9Н	H7	H8	6Н	$HI\theta$	HII
Cu	0.502876	0.330391	0.230499	0.252223	0.241124	0.252223 0.241124 0.277939 0.120737 0.242437	0.120737	0.242437	0.2947	0.273743	0.339967
Mn	0.158482	0.172905	0.172905 0.186172	0.278839	0.242585	0.265306	0.095757	0.278839 0.242585 0.265306 0.095757 0.178942	0.442049 0.263967	0.263967	0.363146
Zn	0.640023	0.451535	0.451535 0.301422	0.481631	0.686838	0.481631 0.686838 0.581145	0.22482	$0.22482 \qquad 0.392517 0.574664 0.449721$	0.574664	0.449721	0.819011
Ba	0	0	0.088654	0.088654 0.139419 0.029227 0.101069	0.029227	0.101069	0.06245		0.080812 0.279965 0.117319	0.117319	0.007727

 Table S3
 Abundance of heavy metals related genes in stool samples from coastal and highland population of Indonesia

Heavy metals					Coast	Coastal nonulation samples	selumbs				
remina Semes	CI	C2	C3	C4	C5	C7	CS CS	62	CIO		
znuC	1826.65	52678.53	1087.99	14308.77	1734.78	470.15	3428.32	7246.33	1039.08		
nikE	3422.41	39377.25	254.81	5828.04	3656.59	0	5709.61	5656.98	486.71		
modC	2608.02	55922.18	776.7	8882.4	1548.04	0	3535.15	10346.03	618.15		
Htum	2229.13	62453.69	98.899	8351.17	1587.77	0	6043.13	69'0002	317.01		
arsB	3746.75	68622.37	0	15677.43	4320.82	1102.11	7813.37	13447.83	101.49		
copR	0	0	273.08	0	0	0	0	0	0		
					Highla	Highland population samples	n samples				
	IH	H2	Н3	H4	H5	9Н	<i>LH</i>	<i>H8</i>	6H	HI0	HII
znuC	13667.83	25642.11	7017.03	55841.14	5185.22	1773.5	24307.83	6054.09	3509.37	2833	0
nikE	13901.56	27536.96	4045.27	42538.33	4857.53	4799.66	19864.63	2835.74	6164.22	3412.23	350.34
modC	17562.97	29913.72	4623.99	59330.4	3172.83	2672.81	19568.16	4321.9	6889.49	4044.85	266.98
Htum	12628.68	25186.29	2305.46	51072.67	4519.81	3967.84	20827.77	5541.03	4550.28	1975.55	228.19
arsB	20597.1	39218.02	8857.27	62324.38	3472.89	3810.97	27581.75	1182.66	11825.75	2608.99	0
copR	0	0	0	0	0	0	0	0	0	0	0