

Asian Journal of Green Chemistry



Original Research Article

View Article Online

View Iournal

Metabolite Profiling of Ethanol Extract of Radix *Uvaria rufa* Blume by UPLC-QToF-MS/MS and Its Potential Aphrodisiac Activity

Maximus M. Taek ¹, Burhan Ma'arif ², * , Dian Nurmawati ³, Novia Maulina ², Diva Dahayu ², Muhammad Alauddin Isyrafi ², Gita Firli Rahmawati ², Faisal Akhmal Muslikh ⁴

ARTICLE INFORMATION

Submitted: 2025-06-02 Revised: 2025-07-08 Accepted: 2025-08-13 Published: 2025-08-17

Manuscript ID: AJGC-2505-1737 DOI: 10.48309/AJGC.2025.520649.1737

KEYWORDS

Metabolite profiling Uvaria rufa blume UPLC-QToF-MS/MS Aphrodisiacs

ABSTRACT

Uvaria rufa Blume, locally called Lelak, has been traditionally used by people in East Nusa Tenggara (NTT), Indonesia, as an herbal medicine to treat erectile dysfunction. Despite its relevance in ethnomedicine, comprehensive scientific data on the metabolite profiling of *U. rufa* radix is still limited. This study aims to determine the metabolite profile of 70% ethanol extract of U. rufa radix using the Ultra Performance Liquid Chromatography-Quadrupole-Time-of-Flight-Mass Spectrometry (UPLC-QToF-MS/MS) method and to determine compounds predicted as aphrodisiacs based on literature studies and preliminary result in silico studies using the SuperPred website. A total of 48 metabolites were identified, with 7-Methoxy-2H-1,3-benzodioxole-5-carboxylic acid (22.14%), Formononetin (15.57%), and 3',7-Dimethoxy-3-hydroxyflavone (14.38%) being the most abundant compounds. Review of relevant literature shows that several identified compounds including Formononetin, 3',7-Dimethoxy-3hydroxyflavone, and Boldine are known to have important biological activities, particularly antioxidant properties. Additionally, prediction results obtained through the SuperPred website indicate that five metabolites may exhibit potential interactions with molecular targets associated with erectile dysfunction. Collectively, these findings provide initial scientific support for the traditional use of U. rufa radix and provide a basis for further pharmacological studies on its prospective aphrodisiac activity.

© 2025 by SPC (Sami Publishing Company), Asian Journal of Green Chemistry, Reproduction is permitted for noncommercial purposes.

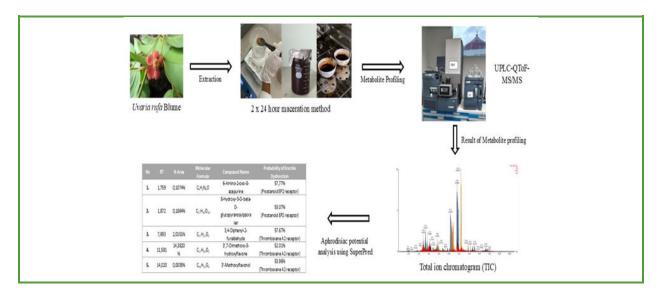
¹Department of Chemistry, Faculty of Science and Technology, Widya Mandira Catholic University, Kupang, Indonesia

²Department of Pharmacy, Faculty of Medicine and Health Science, Maulana Malik Ibrahim State Islamic University, Malang, East Java, Indonesia

 $^{^3}$ Department of Pharmacy, Faculty of Health Science, Kadiri University, Kediri, Indonesia

⁴Department of Pharmacy, Faculty of Pharmacy, Hang Tuah University, Surabaya, Indonesia

Graphical Abstract



Introduction

Indonesia is a tropical country with diverse climates and topography, contributing to its rich and unique biodiversity. Its tropical forests cover approximately 143 million hectares, and around 80% of the world's 28,000 medicinal plant species originate from Indonesia. This biodiversity includes various endemic flora and fauna adapted to their respective habitats [1,2]. Approximately 100 to 150 plant families are distributed across Indonesia, most of which have the potential to be utilized in various industries as sources of fruits, spices, and medicinal raw materials [3]. Additionally, based on empirical data and traditional practices passed down through generations, Indonesians have long utilized plants and animals as alternative medicines for various diseases [4]. Uvaria rufa Blume, commonly known as Lelak, is a species of liana belonging to the Annonaceae family. These plants are distributed across tropical to subtropical regions at altitudes of up to 1500 meters above sea level. Their distribution includes Indonesia, China, Laos, Thailand, Malaysia, Papua New Guinea, the Philippines, Singapore, Vietnam, and India

[5]. In Indonesia, one of the traditional uses of this plant can be found in the East Nusa Tenggara (NTT) region. In this area, U. rufa is locally known as Lekem and thrives in lowland forests as well as in the mountainous areas of Ruteng. The local community utilizes its bark and roots for traditional medicinal purposes [6]. Additionally, this plant is incorporated into herbal concoctions aimed at maintaining physical fitness, preventing illness, restoring vitality and stamina after sickness including enhancing male stamina. generations, people have prepared decoctions from the cortex, stems, and roots of *U. rufa* for consumption [7,8]. Previous research reported that *U. rufa* contains various secondary metabolite compounds, such as flavonoids, terpenoids, steroids, aromatic compounds, and alkaloids [9]. Metabolite profiling is an analytical technique within the metabolomics approach that is used to characterize the profiles of secondary metabolites in plants [10]. This method provides a comprehensive overview of metabolite composition and enables the simultaneous detection of multiple compounds, providing valuable insights for future research.

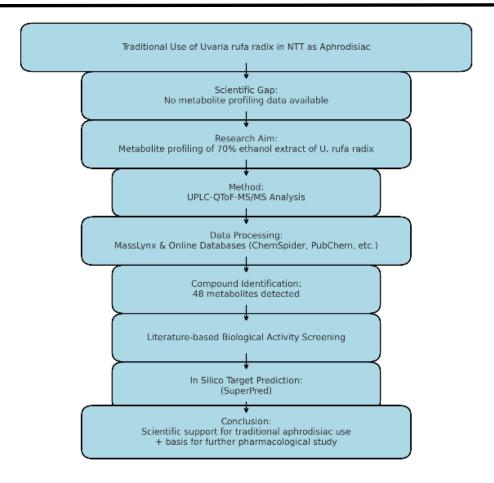


Figure 1. Overview of the research workflow

Such identification has become a widely adopted strategy for revealing secondary metabolite profiles without the need for extensive metabolite isolation processes [11]. The overall experimental workflow of this study is illustrated in Figure 1.

The identification of secondary metabolites in plants can be conducted using UPLC-QToF-MS/MS instruments. This instrument integrates two different methods UPLC and QToF-MS/MS which function to map metabolite profiles, mass spectra, and chromatograms. UPLC-QToF-MS/MS generates qualitative data in the form of metabolite profiles through chromatogram peaks and quantitative data by measuring the percentage concentration of each compound based on chromatogram peak areas [12].

In view of the lack of comprehensive scientific data on the secondary metabolites of U. rufa radix and its traditional use as an aphrodisiac in NTT this study aimed to profile the metabolites of a 70% ethanol extract of U. using rufa radix UPLC-QToF-MS/MS. Furthermore, potential bioactive compounds associated with aphrodisiac activity were identified through a literature review and predictive pharmacological analysis. findings are expected to provide a scientific rationale for its traditional application and to pharmacological support future particularly in the management of erectile dysfunction.

Experimental

Materials

The root of *U. rufa* were collected from Kupang, East Nusa Tenggara, Indonesia, and identified at the Phytochemistry Laboratory, Faculty of Pharmacy, Widya Mandira University, Kupang with number 234/WM.H9/KET/XI/2024. The chemicals used study included 70% dichloromethane, methanol, acetonitrile, and formic acid (all from Merck, Germany). Sterile distilled water (aquadest) was prepared in the laboratory. Instruments used in this study consisted of an ultrasonic cleaner (Sonica, Italy), a rotary evaporator (Heidolph, Germany), and an ACQUITY UPLC® H-Class System coupled with a Xevo G2-S QToF Mass Spectrometer (Waters Corp., USA). The UPLC-QToF-MS/MS analysis was conducted at the Forensic Laboratory, Indonesian Police Criminal Investigation Agency, East Jakarta.

Methods

Plant material and authentication

The roots of *Uvaria rufa* were collected from Kupang, East Nusa Tenggara, Indonesia. The plant material was authenticated at the Phytochemistry Laboratory, Faculty of Pharmacy, Widya Mandira University, Kupang. A voucher specimen was deposited under the registration number 234/WM.H9/KET/XI/2024.

Extraction

The *U. rufa* radix dried plant material extract was obtained using the maceration method. The dried plant material was soaked in a solvent at a ratio of 1:15. During the first maceration, it was immersed in nine parts of the solvent, stirred

for 30 min, and subsequently left to stand for 24 h at ambient temperature in a container covered with aluminum foil. The second maceration was performed by adding six parts of the solvent and allowing it to stand for another 24 h. The macerated filtrate was then filtered and concentrated using a rotary evaporator at 50 °C, with a pressure of 175 psi and a rotation speed of 70 rpm, until the solvent volume significantly reduced. was remaining solvent was further evaporated in an oven at 50 °C to obtain a thick extract. Once the dry extract was obtained, the percentage yield was calculated as the final step. The yield was determined using Equation (1) [13]:

$$Yield (\%) = \frac{Weight \ of \ Extract}{Weight \ of \ dried \ plant \ material} \times 100\% \ (1)$$

Metabolite profiling

The metabolite profiling process was conducted using UPLC-QToF-MS/MS instrument. The steps taken were a 70% ethanol extract of *U. rufa* radix as much as 10 mg dissolved with 10 ml of solvent and inserted into the conditioned Solid Phase Extraction (SPE) column. An aliquot of 5 µL of the prepared sample was injected into the UPLC system using a micro syringe. Chromatographic separation was performed on an ACUITY UPLC® HSS C18 column (1.8 μ m, 2.1 \times 150 mm; Waters). The mobile phases consisted of (A) water with 0.1% formic acid and (B) acetonitrile with 0.1% formic acid. The elution was conducted using a gradient system at a flow rate of 0.2 mL/min for 23 minutes. The separation was conducted under reversed-phase conditions, using a nonpolar stationary phase (C18 column) and polar mobile phases. Mass spectrometric analysis was performed using a Xevo G2-S QToF mass spectrometer (Waters, USA) equipped with an electrospray ionization (ESI) source operating in positive ion mode. The instrument was

operated in Quadrupole followed by Time-of-Flight (ToF) mode, with a collision energy of 4 V and 25–70 V, a mass analysis range of m/z 30–1200, a source temperature of 100 °C, a desolation temperature of 350 °C, and a desolation gas flow of 793 L/h.

Data identification and result interpretation

The raw data from the analysis, in the form of chromatograms and spectra, were further analyzed using MassLynx 4.1 software to obtain information on peak area, retention time, measured mass, calculated mass, and the molecular formula of each detected compound [4]. The confirmation of compound formulas was performed using online database websites such as ChemSpider, MassBank, and PubChem. The selection and confirmation of compounds in the database were based on the highest number of relevant publications and the obtained spectra. The probability of the predicted compounds with activity against erectile determined dysfunction was using SuperPred website by entering the Simplified Molecular Input Line Entry System code (SMILES).

Results and Discussion

The extraction of *U. rufa* radix in this study was conducted using the maceration method. Maceration is an extraction technique that isolates filtrates from solid samples using specific solvents. This method operates based on the "like dissolves like" principle, where the solvent penetrates plant cell walls and saturates intracellular spaces. The concentration gradient between intracellular and extracellular environments facilitates the diffusion of active compounds into the solvent [14]. The final obtained extract weighed 22.4 g, while the initial weight of dried plant material was 309 g,

resulting in a 7.2% yield of the 70% ethanol extract of *U. rufa* radix (Table 1).

Table 1. Yield of 70% ethanol extract of *U. rufa* radix

Dried Plant Material (g)	Extract (g)	Yield (%)
309	22.4	7.2

Metabolite profiling is a type of analysis that utilizes a metabolomics approach to describe or map the profile of secondary metabolite compounds in plants [10].Based phytochemical studies, ethanol extracts of Uvaria rufa roots contain flavonoids, alkaloids, highly oxidized cyclohexane, rutin, isoquercetin, 3-0-β-D-galactopyranoside, kaempferol astragalin, isoquercitrin-6-acetate, benzoylated derivatives, flavonols, kaempferol, quercetin, and lignan glycosides [15]. Prior to injection into the instrument, the sample was pretreated with methanol using the SPE method. The purpose of SPE in the 70% ethanol extract of *U.* rufa radix is to enhance sensitivity and sample analysis. selectivity in enhancing the efficiency of analyte separation from the matrix [16]. Methanol with its high polarity index is able to attract polar secondary metabolites such as flavonoid glycosides, tannins, and some alkaloids. Additionally, the solvent is effective for phenolic compounds with low molecular weight and intermediate polarity [17]. The analysis process in this study employed a reversed-phase system, where the stationary phase is nonpolar, and the mobile reversed-phase phase is polar. In chromatography, more polar substances elute first and have shorter Retention time (Rt) compared to nonpolar substances [18]. The C18 stationary phase, consisting of an 18-carbon hydrocarbon chain, provides strong hydrophobic interactions, making it highly effective for retaining nonpolar or slightly polar analytes [19].

processing and interpretation of chromatogram and spectral data performed using MassLynx 4.1 software, which designed to specifically analyze the chromatographic patterns and spectral characteristics of each detected peak. This software facilitates the identification and prediction of the molecular formula of each compound present in the sample. chromatogram interpretation results include chromatographic peaks, retention times, and m/z values [20]. The confirmation of compound formulas was conducted using online databases such as ChemSpider, MassBank, and PubChem. The selection and validation of compounds from these databases were based on the number of relevant publications and the spectral data obtained [18]. The identification of metabolite profile compounds was further supported by literature studies. Literature searches were conducted using secondary sources that discuss compounds found in *U. rufa* and their associated metabolite profiles in the 70% ethanol extract of *U. rufa* radix. The retrieved data were then classified according to the research formula, processed through reference citation, and compiled into structured abstractions to obtain comprehensive data. These data were subsequently interpreted to generate knowledge that supports the formulation of conclusions. The results of the Total Ion Chromatogram (TIC) and the compounds analyzed from *U. rufa* radix extract with methanol preparation are presented in Figure 2 and Table 2.

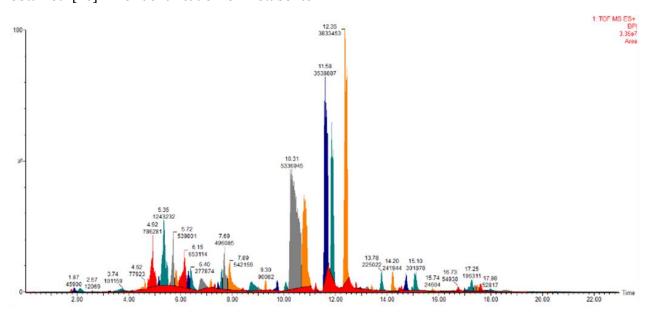


Figure 2. Total ion chromatogram (TIC) of 70% ethanol extract of *U. rufa* Radix

Table 2. Predicted compounds in 70% ethanol extract of *U. rufa* with methanol solvent

	n=		Measured	compouna Calculate	S In 70% etr Molecular	anol extract of <i>U. ruf</i>		
No.	RT	% Area	Mass	d Mass	Formula	Compound Name	Structure	Activity
1	1.759	0.1074 %	150.0286	150.0290	C ₄ H ₂ N ₆ O	6-Amino-2-oxo-8- azapurine	NH ₂	Antiplatelet [21]
2	1.872	0.1864 %	380,0807	380.0728	C ₁₇ H ₁₆ O ₁₀	8-Hydroxy-5-O-beta- D- glucopyranosylpsorale n	110 O O O O O O O O O O O O O O O O O O	-
3	2.110	0.3080 %	461.1693	461.1695	C ₂₃ H ₂₇ NO ₉	Morphine-3-glucuronide	100	-
4	2.575	0.0490 %	267.0976	267.0981	$C_{10}H_{13}N_5O_4$	Rhamnopterin		-
5	2.792	0.0267 %	120.0579	120.0575	C ₈ H ₈ O	Acetophenone		Antioxidant and anti-inflammatory [22,23]
6	2.989	0.0073 %	309.1214	309.1213	C ₁₅ H ₁₉ NO ₆	3-Benzo [1,3] dioxol- 5-yl-3-tert- butoxycarbonylamino- propionic acid		-
7	3.257	0.0396 %	159.0687	159.0684	C ₁₀ H ₉ NO	3-Acetylindole		Antibacterial, anti- HIV, and anti- inflammatory [24,25]
8	3.742	0.4109 %	Unknown	Unknown	Unknown	Unknown	-	-
9	4.136	0.0400 %	193.1110	193.1110	C ₄ H ₁₅ N ₇ S	Unknown	-	-
10	4.621	0.3165 %	471.1168	471.1166	$C_{23}H_{21}NO_{10}$	N-{{[2-(2,5- Dimethoxybenzyliden e)-3-oxo-2,3-dihydro- 1-benzofuran-6-yl] oxy} acetyl) aspartic acid	19430-X.	-
11	4.923	3.4558 %	313.1318	313.1314	C ₁₈ H ₁₉ NO ₄	N-feruloyltyramine		Antioxidant, (Reduces ROS levels), and antidiabetic [26-28]
12	5.345	5.0500 %	327.1477	327.1478	$C_{12}H_{21}N_7O_2$	Boldine	NO CO	Antioxidant, neuroprotective, analgesic, and anticancer [29]
13	5.718	2.1927 %	313.1691	313.1688	C ₁₅ H ₁₉ N ₇ O	3-amino-N-[4-[(3S)-3-amino-1-piperidyl]-3-pyridyl] pyrazine-2-carboxamide		-
14	5.801	0.6892 %	313.1686	357.1955	C ₂₁ H ₂₇ NO ₄	1-(3,4- Dimethoxybenzyl)- 6,7-dimethoxy-2- methyl-1,2,3,4-		-

No.	RT	% Area	Measured Mass	Calculate d Mass	Molecular Formula	Compound Name	Structure	Activity
						tetrahydroisoquinolin e		
15	6.153	2.6529 %	323.1168	323.1171	$C_{20}H_{13}N_5$	3-[5-(2-Naphthyl)-1H- tetrazol-1-yl] quinoline		-
16	6.287	0.8211 %	309.1008	309.1013	$C_{19}H_{11}N_5$	1-Amino-3- phenylpyrido[1,2-a] benzimidazole-2,4- dicarbonitrile		-
17	6.400	1.1287 %	313.1686	341.1682	$C_{20}H_{23}NO_4$	Thaliporphine	10	Antioxidant and anti-inflammatory [30]
18	6.793	1.1985 %	291.0534	291.0532	C ₁₇ H ₉ NO ₄	10- Hydroxyliriodenine	MO	-
19	7.166	0.3661 %	337.1314	337.1314	$C_{20}H_{19}NO_4$	Tetrahydrocorysamin e		Anti-cancer [31]
20	7.320	0.1571 %	352.1425	352.1428	$C_{20}H_{20}N_2O_4$	Feruloylserotonin		-
21	7.454	0.2332 %	309.1365	309.1365	$C_{19}H_{19}NO_3$	Laureline		-
22	7.588	0.8181 %	313.1319	313.1314	C ₁₈ H ₁₉ NO ₄	Laurolistine	NO PORTOR OF THE	-
23	7.693	2.0151 %	248.0843	248.0838	$C_{17}H_{12}O_2$	3,4-Diphenyl-2- furaldehyde		-
24	7.890	2.2022 %	278.0954	278.0950	$C_{18}H_{14}O_3$	Dihydrotanshinone I		-
25	8.396	0.1137 %	309.1359	309.1359	C ₁₉ H ₁₉ NO ₃	Koenigine	10	-
26	8.551	0.0202 %	339.1471	339.1471	$C_{20}H_{21}NO_4$	Papaverine		Pulmonary vasoconstriction, anti-viral, anti- inflammatory, and antiviral [32,33]
27	8.747	1.0114	305.0692	305.0688	$C_{18}H_{11}NO_4$	Oxolaureline		-

No.	RT	% Area	Measured Mass	Calculate d Mass	Molecular Formula	Compound Name	Structure	Activity
28	9.015	0.1943 %	337.1314	337.1314	$C_{20}H_{19}NO_4$	Dihydroberberine		Antidiabetic (increase in insulin) [34]
29	9.296	0.3659 %	366.1103	366.1104	C ₂₁ H ₁₈ O ₆	Glycyrol	NO-00-01	Immunomodulato r of inflammatory arthritis [35]
30	9.450	0.0358 %	297.1369	297.1365	C ₁₈ H ₁₉ NO ₃	Oripavine		Analgesics [36]
31	10.07 0	0.4188 %	284.0689	284.0685	$C_{16}H_{12}O_5$	Wogonin	HO OH O	Antioxidant, anti- inflammatory, and anticancer [37,38]
32	10.30 8	22.143 9%	196.0379	196.0381	C9H8O5	7-Methoxy-2H-1,3- benzodioxole-5- carboxylic acid	NO O	-
33	10.77	9.1982 %	166.0270	166.0266	C ₈ H ₆ O ₄	Phthalic acid	ОН	Anticancer, immunomodulator y, antimicrobial, and insecticidal activity [39,40]
34	11.22 9	0.3207 %	304.0953	304.0952	$C_{16}H_{16}O_{6}$	Oxypeucedanin		Antiproliferative [41]
35	11.58 1	14.382 0%	298.0846	298.0841	C ₁₇ H ₁₄ O ₅	3',7-Dimethoxy-3- hydroxyflavone		Antioxidant and anti-invasive [42-44]
36	11.84 8	8.1381 %	298.0847	298.0846	$C_{17}H_{14}O_5$	3',4'- Dimethoxyflavonol	\$\frac{1}{2} \\ \frac{1}{2} \\ \frac	Neuroprotective [45]
37	12.35 5	15.571 4%	268.0739	268.0736	$C_{16}H_{12}O_4$	Formononetin		Antioxidant, anti- inflammatory, anti-tumour, and agent that enhances adipocyte thermogenesis [46,47]
38	12.79 0	0.3223 %	408.1208	407.1214	$C_{23}H_{20}O_7$	6a,12a- Didehydroamorphigen in		-
39	12.96 6	0.0665 %	374.1153	374.1157	C ₂₃ H ₁₈ O ₅	methyl 4-{[(4-methyl-6-oxo-6H-benzo[c]chromen-3-yl) oxy] methyl} benzoate		

No.	RT	% Area	Measured Mass	Calculate d Mass	Molecular Formula	Compound Name	Structure	Activity
40	13.05 8	0.0062 %	268.0740	268.0736	$C_{16}H_{12}O_4$	Dalbergin	HO	Antifungal, antidiabetic, and antiosteoporotic [48]
41	13.23 3	0.1362 %	409.1471	409.1467	C ₃₀ H ₁₉ NO	2,3,4-Triphenyl-5H-indeno[1,2-b] pyridin-5-one		-
42	13.51 4	0.0221 %	117.0582	117.0579	C ₈ H ₇ N	Indole		-
43	13.58 5	0.0243 %	495.3325	495.3322	$C_{27}H_{41}N_7O_2$	5-({4- [Butyl(hexyl)amino]- 6-(pentylamino)- 1,3,5-triazin-2-yl} amino)-2-methyl-1H- isoindole-1,3(2H)- dione		-
44	13.78 2	0.9140 %	993.6625	993.6627	C53H87N9O9	Unknown		-
45	14.02 0	0.0035 %	268.0741	268.0736	$C_{16}H_{12}O_4$	3'-Methoxyflavonol	HO OH	-
46	14.19 6	0.9828 %	108.3661	108.3661	C ₅₈ H ₉₆ N ₁₀ O	Unknown		-
47	14.54 8	0.3120 %	450.2039	450.2036	$C_{19}H_{34}N_2O_8$ S	5,7-Dimethoxy-3',4'-diprenyloxyisoflavone		-
48	14.72 3	0.8230 %	450.2039	450.2043	C ₂₇ H ₃₀ O ₆	3,5,6-Tris(benzyloxy)- 1,2,4-cyclohexanetriol	OH 25-	-

compound identification results revealed 44 known compounds and 4 unknown compounds based on a database literature The unknown compounds were detected at retention times (RT) of 3.742, 4.136, 13.782, and 14.196 minutes. These unknown compounds may represent impurity compounds that were still detectable by the method or potentially new compounds that have not yet been registered in online databases such as ChemSpider, particularly if their concentrations are high. The results of the analysis obtained several major compounds or dominant compounds identified from data interpretation in Table 2. Major compounds are compounds that have higher levels, as indicated by the percentage of area, compared to other compounds contained in the extract [10]. The major compound in the 70% ethanol extract of U. rufa radix with preparation using methanol is the compound 7-Methoxy-2H-1,3-benzodioxole-5-carboxylic acid with a percentage area of 22.1439%, while several compounds with the highest percentage area are Formononetin having a percentage area of 15.5714% and 3',7-Dimethoxy-3-hydroxyflavone with a percentage area of 14.3820%. There has been no research the compound 7-Methoxy-2H-1,3on benzodioxole-5-carboxylic acid. Meanwhile, formononetin is an isoflavone belonging to the

phytoestrogen group [49,50]. As a result of the literature search, there are several compounds with biological activity. The major compound, 7-Methoxy-2H-1,3-benzodioxole-5-carboxylic acid, is a derivative of piperonylic acid, which is known to have various biological activities. Piperonylic acid has been reported to have anticancer, antioxidant, and antibacterial activities [51-54]. Formononetin compounds have activities as antioxidants, antiinflammatory [46], antitumor, and agents that increase adipocyte thermogenesis [47]. A study explains that formononetin has an antioxidant mechanism by reducing Reactive Oxygen Species (ROS) produced due to exposure to H₂O₂, as well as stabilizing the redox conditions of cells [55]. The compound 3',7-Dimethoxy-3hydroxyflavone has biological activity as an antioxidant that protects cells from free radical damage [41,42], as well as an anti-invasive that plays a role in inhibiting the spread of invasive cells, such as in cancer cases [44]. Additionally, the compounds identified in the 70% ethanol extract of *U. rufa* radix also have several activities, including: Phthalic acid compound with an area percentage of 9.1982% has anticancer, immunomodulatory [39], and antimicrobial activity [40], Boldine compound with an area percentage of 5.0500% has antioxidant and neuroprotective activity [29], N-feruloyltyramine compound with an area percentage of 3.4558% has antioxidant and antidiabetic activity [28], Wogonin compound with an area percentage of 0.4188% has antiinflammatory and anticancer activity [37,38], and Dihydroberberine compound with an area percentage of 0.1943% has antidiabetic activity by increasing insulin [34]. The presence of antioxidants is essential in counteracting oxidative stress and maintaining normal penile function. Compounds such as superoxide dismutase (SOD), vitamin C, vitamin E, melatonin, alpha-lipoic acid, peroxynitrite decomposition catalysts, and gamma-linolenic acid have been recognized for their ability to minimize diabetic vasculopathy and autonomic neuropathy affecting the penile tissue, which in turn contributes to the improvement of erectile performance at various physiological level [42,56-61]. Evidence from animal studies involving arteriogenic erectile dysfunction models has indicated that pomegranate juice may reduce oxidative stress and enhance erectile function [56]. These favorable effects are largely associated with the high content of antioxidants polyphenolic present pomegranate, which are believed to support vascular integrity and erectile capacity [62]. Antioxidant activity plays a crucial role in erectile dysfunction treatment by reducing oxidative stress caused by Reactive Oxygen Species (ROS) and enhancing blood flow to erectile tissue, thereby offering therapeutic potential [63]. Although the antioxidant activity in several secondary metabolites of 70% ethanol extract of Uvaria rufa radix has not been proven to directly improve erectile function, the presence of compounds related to antioxidants indicates the potential that can be developed and analyzed more deeply regarding the therapeutic effects of erectile dysfunction. There are five compounds identified in the 70% ethanol extract of *U. rufa* radix, namely 6-Amino-2-oxo-8-azapurine, 8-Hydroxy-5-0-beta-D-glucopyranosylpsoralen, 3,4-Diphenyl-2furaldehyde, 3',7-Dimethoxy-3-hydroxyflavone, and 3'-Methoxyflavonol. Several of these compounds were simulated using the SuperPred website regarding the percentage of probability of indications of erectile dysfunction. The website uses the principle of similarity of physicochemical properties and biological activity.

Table 3. Predicted 1	probability of com	pounds against	erectile dysfunction
rable of realestea	probability of com	pourido agarriot	creeine aybranetion

No.	RT	% Area	Molecular Formula	Compound Name	Probability of Erectile Dysfunction
1.	1.759	0.1074%	$C_4H_2N_6O$	6-Amino-2-oxo-8- azapurine	57,77% (Prostanoid EP2 receptor)
2.	1.872	0.1864%	$C_{17}H_{16}O_{10}$	8-Hydroxy-5-0-beta-D- glucopyranosylpsoralen	53.07% (Prostanoid EP2 receptor)
3.	7.693	2.0151%	C ₁₇ H ₁₂ O ₂	3,4-Diphenyl-2- furaldehyde	57.67% (Thromboxane A2 receptor)
4.	11.581	14.3820%	C ₁₇ H ₁₄ O ₅	3',7-Dimethoxy-3- hydroxyflavone	52.01% (Thromboxane A2 receptor)
5.	14.020	0.0035%	$C_{16}H_{12}O_4$	3'-Methoxyflavonol	53.96% (Thromboxane A2 receptor)

Its use can be done by uploading a molecular structure or entering the SMILE code, then the system will simulate the reaction of compound and protein interactions with the data in the database [64]. The SuperPred analysis results Table 3 identified five compounds with a probability percentage indicating their potential relevance to erectile dysfunction. Several receptors were found to predict the probability of compounds against erectile dysfunction, Prostanoid namelv EP2 receptor Thromboxane A2 receptor [65,66]. The EP2 receptor is a member of the G protein-coupled receptor (GPCR) family involved in the prostanoid signaling pathway. This receptor regulates blood flow to erectile tissue by activating Gs protein-mediated signaling, which increases the production of cyclic adenosine monophosphate (cAMP). Elevated cAMP levels activate protein kinase A (PKA), leading to a reduction in intracellular calcium levels and promoting smooth muscle relaxation in the corpus cavernosum, thereby facilitating penile erection [67,68]. Thromboxane A2 receptor (TP) is a receptor that belongs to the G proteinbound receptor (GPCR) group. This receptor plays a role in various physiological and pathophysiological processes, such as platelet aggregation and smooth muscle contraction. Thromboxane A2 (TXA2) receptor binding to TP

receptors can trigger smooth muscle contraction in penile arteries through the activation of Rho kinase (ROCK). If TXA2 and the ROCK pathway are inhibited, smooth muscle contraction will be reduced. This causes the smooth muscle in the penile arteries to relax, resulting in increased blood flow to the erectile tissue [69,70]. The percentage probability of the compound 3',7-Dimethoxy-3-hydroxyflavone is 52.01% with a possible target receptor TXA2. The compound has a percentage area of 14.3820%, and could be a potential compound responsible for the therapeutic effect of *U. rufa* radix. The compound 3',7-Dimethoxy-3hydroxyflavone is included in the secondary metabolites of the flavonoid group. Flavonoid compounds can be antagonists for TXA2 by reducing the mechanism of platelet aggregation induced by TXA2 agonists [71]. The percentage of probability generated by the SuperPred website is still a prediction based on simulations from the website's machine learning model based on available databases such as CheEMBL. The percentage generated from the 5 compounds is still relatively low, and further confirmation must be done [64,72]. This confirmation can be done through in silico analysis using a molecular docking approach to obtain more accurate parameters as a reference for the next test. High probability results, such

as (>80%) for the interaction of a compound and a biological target, indicate high confidence in predictions based on simulated data patterns. High probabilities support the belief that the compound interacts with the target, while low probabilities indicate the need for further validation [72].

Conclusion

This study successfully characterized the metabolite profile of a 70% ethanol extract of Uvaria rufa radix using UPLC-QToF-MS/MS, resulting in the identification of 48 secondary metabolites. The major compounds identified 7-methoxy-2H-1,3-benzodioxole-5included carboxylic acid, Formononetin, and 3',7dimethoxy-3-hydroxyflavone. Literature based evidence suggests that several of these compounds possess biological properties, particularly antioxidant activity. Furthermore, target prediction through the SuperPred platform indicated potential interactions between five metabolites and targets related to erectile dysfunction. These findings provide preliminary scientific support for the traditional use of *U. rufa* radix and serve as a foundation for future pharmacological and mechanistic studies exploring its potential aphrodisiac effects.

Acknowledgments

The authors would like to express their gratitude to all parties who have contributed to this research, from the data collection process to the analysis. They sincerely appreciate their hard work and dedication in ensuring the success of this study.

Disclosure Statement

The authors declare no conflict of interest.

Funding

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

Authors' Contributions

Conceptualization, Burhan Ma'arif and Maximus M. Taek; Methodology, Burhan Ma'arif, Dian Nurmawati, and Novia Maulina; Investigation, Diva Dahayu, Muhammad A. Isyrafi, and Gita F. Rahmawati; Data curation, Diva Dahayu, Muhammad A. Isyrafi, and Gita F. Rahmawati; Formal analysis, Maximus M. Taek and Burhan Ma'arif; Writing—original draft preparation, Burhan Ma'arif and Maximus M. Taek; Writing—review and editing, Faisal A. Muslikh and Burhan Ma'arif; Supervision and Project administration, Burhan Ma'arif; All authors have read and agreed to the published version of the manuscript.

Orcid

Maximus M. Taek https://orcid.org/0000-0002-4597-2167 Burhan Ma'arif https://orcid.org/0000-0001-9182-343X Dian Nurmawati https://orcid.org/0000-0003-0750-0545 Novia Maulina https://orcid.org/0000-0002-7948-0101 Diva Dahayu https://orcid.org/0009-0001-4544-6693 Muhammad A. Isyrafi https://orcid.org/0009-0008-9634-0849 Gita F. Rahmawati https://orcid.org/0009-0006-3572-5249 Faisal A. Muslikh https://orcid.org/0000-0002-9611-7937

References

[1]. Woerdenbag H.J., Kayser O. Jamu: Indonesian traditional herbal medicine towards

rational phytopharmacological use. *Journal of Herbal Medicine*, 2014, **4**:51 [Crossref], [Google Scholar], [Publisher]

- [2]. Haba F.S., Purnama M.M., Mau A.E. Keanekaragaman jenis dan pemanfaatan tumbuhan obat di hutan penelitian bu'at so'e, kecamatan mollo selatan, kabupaten timor tengah selatan, provinsi nusa tenggara timur. *Wana Lestari*, 2022, **4**:182 [Crossref], [Google Scholar], [Publisher]
- [3]. Korja I., Lestari N.I. Keanekaragaman jenis tumbuhan obat di wisata alam desa kapopo kabupaten sigi biromaru provinsi sulawesi tengah. *Forest Science*, 2019, **17** [Crossref], [Google Scholar], [Publisher]
- [4]. a) Ivantarina D., Nurdiana N., Sujuti H., Wiyasa I.W.A. Metabolite profiling and in silico of four varieties of black soybeans (glycine soja) as candidate for preeclampsia treatment. *Advanced Journal of Chemistry, Section A*, 2025, 8:1344 [Crossref], [Publisher]; b) Hakim A., Muti'ah R., Aprinda R., Suryadinata A., Nasikhatul F. Metabolite profiling bagian akar, batang, daun, dan biji helianthus annuus l. menggunakan UPLC-MS. *Media Pharmaceutica Indonesiana*, 2018, 2:64 [Crossref], [Google Scholar], [Publisher]
- [5]. a) Kuswanti N., Qomariyah N., Purnama E.R., Khaleyla, F. Bruguirea gymnorrhyza leaf extract metabolites: Oral bioavailability and gi absorption predictions. *Journal of Medicinal and Chemical Sciences*, 2024, 7:518 [Crossref], [Google Scholar], [Publisher]; b) Tudla F.A., Aguinaldo A.M., Krohn K., Hussain H., Macabeo A.P.G. Highly oxygenated cyclohexene metabolites from Uvaria rufa. *Biochemical Systematics and Ecology*, 2007, 35:45 [Crossref], [Google Scholar], [Publisher]
- [6]. Iswandono E. Budaya konservasi orang manggarai: Studi kasus di daerah penyangga taman wisata alam ruteng-nusa tenggara timur. *Penerbit Balai Besar Konservasi Sumber Daya*

- Alam, Nusa Tenggara Timur, Kupang, 2018, [Google Scholar], [Publisher]
- [7]. Taek M.M., Bambang P.E., Agil M. Plants used in traditional medicine for treatment of malaria by Tetun ethnic people in West Timor Indonesia. *Asian Pacific Journal of Tropical Medicine*, 2018, **11**:630 [Crossref], [Google Scholar], [Publisher]
- [8]. Taek M.M., Mali S. Plants in ai tahan: traditional medicine of the Tetun ethnic community in West Timor Indonesia. *Proc 7th Annual Basic Science Int Conf. Malang: Brawijaya University: 2017*, 2017, 71 [Google Scholar], [Publisher]
- [9]. a) Gavanaroudi S.B. The role of nutrition in chronic disease prevention and management. *SPC Journal of Medical and Healthcare*, 2025,1:50 [Crossref], [Publisher]; b) Macabeo A.P.G., Tudla F.A., Krohn K., Franzblau S.G. Antitubercular activity of the semi–polar extractives of Uvaria rufa. *Asian Pacific Journal of Tropical Medicine*, 2012, 5:777 [Crossref], [Google Scholar], [Publisher]
- [10]. Ma'arif B., Mutiah R. Profil metabolit berbagai ekstrak daun chrysophyllum cainito L. menggunakan UPLC-QTOF-MS/MS. *Jurnal Tumbuhan Obat Indonesia*, 2019, **12**:10 [Google Scholar], [Publisher]
- [11]. a) Taek M.M., Ma'arif B., Muslikh F.A., Maulina N., Nurmawati D., Dean M., Muntasir M. The aprhrodisiac effect of ethanol extract of *Uvaria Rufa* blume. bark on male mice (*Mus Musculus*). *Advanced Journal of Chemistry, Section A*, 2025, **8**:1661 [Crossref], [Publisher]; b) Nurmaida N., Darusman L.K., Rafi M., Heryanto R. Metabolite profiling of tabat barito (ficus deltoidea) using UPLC-QTOF-MS/MS. *The Journal of Pure and Applied Chemistry Research*, 2018, **7**:100 [Crossref], [Google Scholar], [Publisher]
- [12]. a) Yuliani Y., Khaleyla F., Anggorowati Rahayu D. Potential of bioactive compound from elephantopus scaber linn. leaf as anti-cancer

through in silico test. *Journal of Medicinal and Chemical Sciences*, 2023, **6**:1773 [Crossref], [Google Scholar], [Publisher]; b) Pavia D.L., Lampman G.M., Kriz G.S., Vyvyan J.R., Introduction to spectroscopy. *Belmont, USA*, 200113 [Google Scholar], [Publisher]

[13]. Widyastuti I., Luthfah H.Z., Hartono Y.I., Islamadina R., Can A.T., Rohman A. Aktivitas antioksidan temulawak (Curcuma xanthorrhiza Roxb.) dan profil pengelompokannya dengan kemometrik antioxidant activity of temulawak (Curcuma xanthorrhiza Roxb.) and its classification with chemometrics. *Indonesian Journal of Chemometrics and Pharmaceutical Analysis*, 2021, 1:28 [Crossref], [Google Scholar], [Publisher]

[14]. Hujjatusnaini N., Indah B., Afitri E., Widyastuti R., Ardiansyah A. Buku referensi ekstraksi. *IAIN Palangka Raya*, 2021 [Google Scholar], [Publisher]

[15]. Buncharoen W., Saenphet K., Saenphet S., Thitaram C. Uvaria rufa Blume attenuates benign prostatic hyperplasia via inhibiting 5α-reductase and enhancing antioxidant status. *Journal of Ethnopharmacology*, 2016, **194**:483 [Crossref], [Google Scholar], [Publisher]

[16]. Rahmatia T.U. Metode SPE (solid phase extraction) sebagai alternatif terbaru dalam analisis dan pemurnian senyawa obat. *Farmaka*, 2016, **14**:151 [Crossref], [Google Scholar], [Publisher]

[17]. Yusnawan E. The effectiveness of polar and non polar fractions of Ageratum conyzoides l. to control peanut rust disease and phytochemical screenings of secondary metabolites. *Jurnal Hama dan Penyakit Tumbuhan Tropika*, 2013, 13:159 [Crossref], [Google Scholar], [Publisher] [18]. Harmita K.A.A., Harahap Y., Supandi. Liquid Chromatography-Tandem Mass Spectrometry (LC-MS/MS). *Jakarta Barat: Isfi Penerbitan*, 2019 [Publisher]

[19]. Lestari P.E.D., Jannah A.K.R., Fitri M.N., Azharani N.A. Analisis Perbandingan HPLC dan

Teknik Lain untuk Deteksi Antibiotik. *Termometer: Jurnal Ilmiah Ilmu Kesehatan dan Kedokteran*, 2024, **2**:01 [Crossref], [Google Scholar], [Publisher]

[20]. Mutiah R., Bhagawan W.S., Ma'arif B., Rahmandika J.M.S. Metabolite Fingerprinting Eleutherine palmifolia (L.) Merr. Using UPLC-QTOF-MS/MS. *Majalah Obat Tradisional*, 2019, **24**:139 [Crossref], [Google Scholar], [Publisher] [21]. Zhao Z., Wang Y., Tian N., Yan H., Wang J. Synthesis and biological evaluation of N 6 derivatives of 8-azapurine as novel antiplatelet agents. *RSC Medicinal Chemistry*, 2021, **12**:1414 [Crossref], [Google Scholar], [Publisher]

[22]. Wang A., Gao X., Huo X., Huang S., Feng L., Sun C., Zhang B., Ma X., Jia J., Wang C. Antioxidant acetophenone glycosides from the roots of Euphorbia ebracteolata Hayata. *Natural Product Research*, 2018, **32**:2187 [Crossref], [Google Scholar], [Publisher]

[23]. Chen N.H., Li W., Zhong Y.L., Niu Q.W., Li Y.Y., Zhang Y.B., Li M.M., Li Y.L., Wang G.C. New acetophenone derivatives from acronychia oligophlebia and their anti - inflammatory and antioxidant activities. *Chemistry & Biodiversity*, 2018, **15**:e18000080 [Crossref], [Google Scholar], [Publisher]

[24]. Metwally M.A., Shaaban S., Abdel-Wahab B.F., El-Hiti G.A. 3-Acetylindoles: Synthesis, reactions and biological activities. *Current Organic Chemistry*, 2009, **13**:1475 [Crossref], [Google Scholar], [Publisher]

[25]. Rani P., Srivastava V., Kumar A. Synthesis and antiinflammatory activity of heterocyclic indole derivatives. *European Journal of Medicinal Chemistry*, 2004, **39**:449 [Crossref], [Google Scholar], [Publisher]

[26]. Forino M., Tartaglione L., Dell'Aversano C., Ciminiello P. NMR-based identification of the phenolic profile of fruits of Lycium barbarum (goji berries). Isolation and structural determination of a novel N-feruloyl tyramine dimer as the most abundant antioxidant

polyphenol of goji berries. *Food Chemistry*, 2016, **194**:1254 [Crossref], [Google Scholar], [Publisher]

[27]. Cavin A., Hostettmann K., Dyatmyko W., Potterat O. Antioxidant and lipophilic constituents of Tinospora crispa. *Planta Medica*, 1998, **64**:393 [Crossref], [Google Scholar], [Publisher]

[28]. Amaro C.A.B., González-Cortazar M., Herrera-Ruiz M., Román-Ramos R., Aguilar-Santamaría L., Tortoriello J., Jiménez-Ferrer E. Hypoglycemic and hypotensive activity of a root extract of Smilax aristolochiifolia, standardized on N-trans-feruloyl-tyramine. *Molecules*, 2014, 19:11366 [Crossref], [Google Scholar], [Publisher]

[29]. Sáez J.C., Burrell J.C., Cahill C.M., Cullen D.K., Devi L.A., Gilbert R.J., Graham Z.A., Gurvich V.J., Havton L.A., Iyengar R. Pharmacology of boldine: summary of the field and update on recent advances. *Frontiers in Pharmacology*, 2024, **15**:1427147 [Crossref], [Google Scholar], [Publisher]

[30]. Ku H.C., Lee S.Y., Lee S.S., Su M.J. Thaliporphine, an alkaloid from Neolitsea konishii, exerts antioxidant, anti-inflammatory, and anti-apoptotic responses in guinea pig during cardiovascular collapse in inflammatory disease. *Journal of Functional Foods*, 2016, **26**:57 [Crossref], [Google Scholar], [Publisher] [31]. Zhou J.B., Peng G., Li J., Jia Y.C., Wang J., Nie S., Zhang Q.Y. Anticancer activity of

tetrahydrocorysamine against pancreatic adenocarcinoma cell line PANC-1 in vitro and in vivo. *Tropical Journal of Pharmaceutical Research*, 2016, **15**:141 [Crossref], [Google Scholar], [Publisher]

[32]. Maggi M., Filippi S., Ledda F., Magini A., Forti G. Erectile dysfunction: from biochemical pharmacology to advances in medical therapy. *European Journal of Endocrinology*, 2000, **143**:143 [Crossref], [Google Scholar], [Publisher]

[33]. Ashrafi S., Alam S., Sultana A., Raj A., Emon N.U., Richi F.T., Sharmin T., Moon M., Park M.N., Kim B. Papaverine: a miraculous alkaloid from opium and its multimedicinal application. *Molecules*, 2023, **28**:3149 [Crossref], [Google Scholar], [Publisher]

[34]. Chang C., Roh Y.S., Du M., Kuo Y.C., Zhang Y., Hardy M., Gahler R., Solnier J. Differences in metabolite profiles of dihydroberberine and micellar berberine in CaCO₂ cells and humans—a pilot study. *International Journal of Molecular Sciences*, 2024, **25**:5625 [Crossref], [Google Scholar], [Publisher]

[35]. Fu Y., Zhou H., Wang S., Wei Q. Glycyrol suppresses collagen-induced arthritis by regulating autoimmune and inflammatory responses. *PloS one*, 2014, **9**:e98137 [Crossref], [Google Scholar], [Publisher]

[36]. Gómez-Serranillos M., Palomino O., Carretero E., Villar A. Analytical study and analgesic activity of oripavine from Papaver somniferum L. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 1998, **12**:346 [Crossref], [Google Scholar], [Publisher]

[37]. Tai M.C., Tsang S.Y., Chang L.Y., Xue H. Therapeutic potential of wogonin: A naturally occurring flavonoid. *CNS Drug Reviews*, 2005, **11**:141 [Crossref], [Google Scholar], [Publisher] [38]. Wang L., Zhang D., Wang N., Li S., Tan H.-Y., Feng Y. Polyphenols of Chinese skullcap roots: from chemical profiles to anticancer effects. *RSC Advances*, 2019, **9**:25518 [Crossref], [Google Scholar], [Publisher]

[39]. Save S., Lokhande R., Chowdhary A. Determination of 1, 2-Benzenedicarboxylic acid, bis (2-ethylhexyl) ester from the twigs of Thevetia peruviana as a Colwell Biomarker. *Journal of Innovations in Pharmaceuticals and Biological Sciences*, 2015, **2**:349 [Google Scholar], [Publisher]

[40]. Huang L., Zhu X., Zhou S., Cheng Z., Shi K., Zhang C., Shao H. Phthalic acid esters: Natural sources and biological activities. *Toxins*, 2021, **13**:495 [Crossref], [Google Scholar], [Publisher] [41]. Park S.H., Hong J.Y., Park H.J., Lee S.K., The antiproliferative activity of oxypeucedanin via induction of G₂/M phase cell cycle arrest and p53-dependent MDM₂/p21 expression in human hepatoma cells. *Molecules*, 2020, **25**:501 [Crossref], [Google Scholar], [Publisher]

[42]. Chen S., Li X., Liu X., Wang N., An Q., Ye X.M., Zhao Z.T., Zhao M., Han Y., Ouyang K.H. Investigation of chemical composition, antioxidant activity, and the effects of alfalfa flavonoids on growth performance. *Oxidative Medicine and Cellular Longevity*, 2020, **2020**:8569237 [Crossref], [Google Scholar], [Publisher]

[43]. Park H., Lee E.J., Moon D., Yun H., Cha A., Hwang I., Kim H.S. Discovery of 3, 7-dimethoxyflavone that inhibits liver fibrosis based on dual mechanisms of antioxidant and inhibitor of activated hepatic stellate cell. *Free Radical Biology and Medicine*, 2023, **204**:195 [Crossref], [Google Scholar], [Publisher]

[44]. Parmar V.S., Jain R., Sharma S.K., Vardhan A., Jha A., Taneja P., Singh S., Vyncke B.M., Bracke M.E., Mareel M.M. Anti-invasive activity of 3, 7-dimethoxyflavone in vitro. *Journal of Pharmaceutical Sciences*, 1994, **83**:1217 [Crossref], [Google Scholar], [Publisher]

[45]. Fatokun A., Liu J., Dawson V., Dawson T. Identification through high-throughput screening of 4'-methoxyflavone and 3', 4'-dimethoxyflavone as novel neuroprotective inhibitors of parthanatos. *British journal of pharmacology*, 2013, **169**:1263 [Crossref], [Google Scholar], [Publisher]

[46]. Ding M., Bao Y., Liang H., Zhang X., Li B., Yang R., Zeng N. Potential mechanisms of formononetin against inflammation and oxidative stress: A review. *Frontiers in*

Pharmacology, 2024, **15**:1368765 [Crossref], [Google Scholar], [Publisher]

[47]. Nie T., Zhao S., Mao L., Yang Y., Sun W., Lin X., Liu S., Li K., Sun Y., Li P. The natural compound, formononetin, extracted from Astragalus membranaceus increases adipocyte thermogenesis by modulating PPARy activity. *British Journal of Pharmacology*, 2018, **175**:1439 [Crossref], [Google Scholar], [Publisher]

[48]. Zhou S., Huang G., Chen G. Synthesis and biological activities of drugs for the treatment of osteoporosis. *European Journal of Medicinal Chemistry*, 2020, **197**:112313 [Crossref], [Google Scholar], [Publisher]

[49]. Dutra J.M., Espitia P.J., Batista R.A. Formononetin: Biological effects and uses–A review. *Food Chemistry*, 2021, **359**:129975 [Crossref], [Google Scholar], [Publisher]

[50]. Jiang D., Rasul A., Batool R., Sarfraz I., Hussain G., Mateen Tahir M., Qin T., Selamoglu Z., Ali M., Li J. Potential anticancer properties and mechanisms of action of formononetin. *BioMed research international*, 2019, **2019**:5854315 [Crossref], [Google Scholar], [Publisher]

[51]. Moreira K.G., do Prado T.P., Mendes N.F., de Medeiros Bezerra R., Jara C.P., Melo Lima M.H., de Araujo E.P. Accelerative action of topical piperonylic acid on mice full thickness wound by modulating inflammation and collagen deposition. *PloS One*, 2021, **16**:e0259134 [Crossref], [Google Scholar], [Publisher]

[52]. Zarai Z., Boujelbene E., Salem N.B., Gargouri Y., Sayari A. Antioxidant and antimicrobial activities of various solvent extracts, piperine and piperic acid from Piper nigrum. *LWT-Food Science and Technology*, 2013, **50**:634 [Crossref], [Google Scholar], [Publisher]

[53]. Lee D., Lim J., Woo K.C., Kim K.T. Piperonylic acid stimulates keratinocyte growth

and survival by activating epidermal growth factor receptor (EGFR). *Scientific Reports*, 2018, **8**:162 [Crossref], [Google Scholar], [Publisher]

[54]. Schalk M., Cabello-Hurtado F., Pierrel M.-A.s., Atanassova R., Saindrenan P., Werck-Reichhart D.l. Piperonylic acid, a selective, mechanism-based inactivator of the transcinnamate 4-hydroxylase: a new tool to control the flux of metabolites in the phenylpropanoid pathway. *Plant Physiology*, 1998, **118**:209 [Crossref], [Google Scholar], [Publisher]

[55]. Li Jian L.J., Han Lin H.L., Ma YuFang M.Y., Huang YiFan H.Y. Inhibiting effects of three components of astragalus membranaceus on oxidative stress in chang liver cells. 2015, [Google Scholar], [Publisher]

[56]. Azadzoi K.M., Schulman R.N., Aviram M., Siroky M.B. Oxidative stress in arteriogenic erectile dysfunction: Prophylactic role of antioxidants. *The Journal of urology*, 2005, **174**:386 [Crossref], [Google Scholar], [Publisher]

[57]. Najafabadi B.T., Jafarinia M., Ghamari K., Shokraee K., Tadayyon F., Akhondzadeh S. Vitamin E and ginseng combined supplement for treatment of male erectile dysfunction: a double-blind, placebo-controlled, randomized, clinical trial. *Advances in Integrative Medicine*, 2021, **8**:44 [Crossref], [Google Scholar], [Publisher]

[58]. Jeffrey S., Samraj P.I., Raj B.S. The role of alpha-lipoic acid supplementation in the prevention of diabetes complications: a comprehensive review of clinical trials. *Current Diabetes Reviews*, 2021, **17**:87 [Crossref], [Google Scholar], [Publisher]

[59]. Fu H., Bai X., Le L., Tian D., Gao H., Qi L., Hu K.-p. Eucommia ulmoides oliv. leaf extract dysfunction improves erectile in streptozotocin - induced diabetic rats by endothelial function protecting and ameliorating hypothalamic pituitary Evidence-Based gonadal axis function.

Complementary and Alternative Medicine, 2019, **2019**:1782953 [Crossref], [Google Scholar], [Publisher]

[60]. Sheweita S.A., Meftah A.A., Sheweita M.S., Balbaa M.E. Erectile dysfunction drugs altered the activities of antioxidant enzymes, oxidative stress and the protein expressions of some cytochrome P450 isozymes involved in the steroidogenesis of steroid hormones. *PloS One*, 2020, **15**:e0241509 [Crossref], [Google Scholar], [Publisher]

[61]. Shivavedi N., Tej G.N.V.C., Neogi K., Nayak P.K. Ascorbic acid therapy: A potential strategy against comorbid depression-like behavior in streptozotocin-nicotinamide-induced diabetic rats. *Biomedicine & Pharmacotherapy*, 2019, **109**:351 [Crossref], [Google Scholar], [Publisher]

[62]. Zarfeshany A., Asgary S., Javanmard S.H. Potent health effects of pomegranate. *Advanced Biomedical Research*, 2014, **3**:100 [Crossref], [Google Scholar], [Publisher]

[63]. Kaltsas A., Zikopoulos A., Dimitriadis F., Sheshi D., Politis M., Moustakli E., Symeonidis E.N., Chrisofos M., Sofikitis N., Zachariou A. Oxidative stress and erectile dysfunction: Pathophysiology, impacts, and potential treatments. *Current Issues in Molecular Biology*, 2024, 46:8807 [Crossref], [Google Scholar], [Publisher]

[64]. Dunkel M., Günther S., Ahmed J., Wittig B., Preissner R. SuperPred: drug classification and target prediction. *Nucleic Acids Research*, 2008, **36**:W55 [Crossref], [Google Scholar], [Publisher]

[65]. Sato M., Kawatani M. Characterization of prostaglandin E receptor subtypes involved in the relaxation of rabbit penile corpus cavernosum smooth muscle. *Biomedical Research*, 2004, **25**:237 [Crossref], [Google Scholar], [Publisher]

[66]. Khan M., Thompson C., Sullivan M., Jeremy J., Mikhailidis D., Morgan R. The role of

prostaglandins in the aetiology and treatment of erectile dysfunction. *Prostaglandins, Leukotrienes and Essential Fatty Acids (PLEFA),* 1999, **60**:169 [Crossref], [Google Scholar], [Publisher]

[67]. Ganesh T. Prostanoid receptor EP₂ as a therapeutic target: Miniperspective. *Journal of Medicinal Chemistry*, 2014, **57**:4454 [Crossref], [Google Scholar], [Publisher]

[68]. Norel X., Sugimoto Y., Ozen G., Abdelazeem H., Amgoud Y., Bouhadoun A., Bassiouni W., Mani S., Manikpurage Goepp M., International union of basic and clinical pharmacology. CIX. Differences and similarities between human and rodent prostaglandin E2 receptors (EP1-4) and prostacyclin receptor Specific roles in pathophysiologic conditions. Pharmacological Reviews, 2020, **72**:910 [Crossref], [Google Scholar], [Publisher] [69]. Sopko N.A., Hannan J.L., Bivalacqua T.J. Understanding and targeting the Rho kinase pathway in erectile dysfunction. Nature Reviews Urology, 2014, 11:622 [Crossref], [Google Scholar], [Publisher]

[70]. Grann M., Comerma-Steffensen S., Arcanjo D.D., Simonsen U. Mechanisms involved in

thromboxane A2-induced vasoconstriction of rat intracavernous small penile arteries. Basic & Clinical Pharmacology & Toxicology, 2016, **119**:86 [Crossref], [Google Scholar], [Publisher] [71]. Guerrero J., Lozano M., Castillo J., Benavente-Garcia O., Vicente V., Rivera J. Flavonoids inhibit platelet function through binding to the thromboxane A₂ receptor. *Journal* of Thrombosis and Haemostasis, 2005, 3:369 [Crossref], [Google Scholar], [Publisher] [72]. Gallo K., Goede A., Preissner R., Gohlke B.-O. SuperPred 3.0: Drug classification and target prediction—a machine learning approach. Nucleic Acids Research, **50**:W726 2022,

How to cite this manuscript: M.M. Taek, B. Ma'arif, D. Nurmawati, N. Maulina, D. Dahayu, M.A. Isyrafi, G.F. Rahmawati, F.A. Muslikh. Metabolite profiling of ethanol extract of radix *uvaria rufa* blume by UPLC-QToF-MS/MS and its potential aphrodisiac activity. *Asian Journal of Green Chemistry*, 9(6) 2025, 985-1003.

DOI: 10.48309/AJGC.2025.520649.1737

[Crossref], [Google Scholar], [Publisher]